

STATEMENT OF EFSA

Statement on a request from the European Commission related to an emergency measure notified by France under Article 34 of Regulation (EC) 1829/2003 to prohibit the cultivation of genetically modified maize MON 810¹

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ABSTRACT

Following a request from the European Commission, the European Food Safety Authority (EFSA) evaluated the documentation submitted by France under Article 34 of Regulation (EC) 1829/2003 in support of its request to prohibit the cultivation of genetically modified maize MON 810 in the EU. Neither the scientific publications cited in the documentation submitted by France with relevance to maize MON 810 nor the arguments put forward by France reveal any new information that would invalidate the previous risk assessment conclusions and risk management recommendations made by the EFSA GMO Panel. EFSA considers that the previous GMO Panel risk assessment conclusions and risk management recommendations on maize MON 810 remain valid and applicable. Therefore, EFSA concludes that, based on the documentation submitted by France, there is no specific scientific evidence, in terms of risk to human and animal health or the environment, that would support the adoption of an emergency measure on the cultivation of maize MON 810 under Article 34 of Regulation (EC) 1829/2003.

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KEY WORDS

GMO, maize (*Zea mays*), MON 810, France, emergency measure, environment, Directive 2002/53/EC

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SUMMARY

On 25 March 2014, the European Commission requested the European Food Safety Authority (EFSA) to assess the supporting documentation submitted by France to notify an emergency measure under Article 34 of Regulation (EC) 1829/2003, prohibiting the cultivation of genetically modified maize MON 810 in the EU.

EFSA assessed the documentation supplied by the French Authorities and the scientific publications cited in the French Authorities' report. For each area of concern outlined in the French Authorities' report, EFSA assessed whether any of the scientific publications not previously addressed by EFSA and/or its GMO Panel, or any of the arguments put forward by France, would invalidate the previous GMO Panel conclusions on the safety of maize MON 810. Moreover, EFSA considered the relevance of concerns raised by France in the light of the most recent and relevant scientific data published in the scientific literature.

During its evaluation of the French Authorities' report, EFSA noted that most of the cited scientific publications were addressed previously by EFSA and its GMO Panel in various scientific outputs. These publications were therefore not considered further. For the remaining scientific publications, EFSA has focused its assessment on those aspects that are relevant to maize MON 810.

Neither the scientific publications cited in the French Authorities' report with relevance to maize MON 810 nor the arguments put forward by France reveal any new information that would invalidate the previous risk assessment conclusions and risk management recommendations made by the EFSA GMO Panel. Therefore, EFSA considers that the previous GMO Panel risk assessment conclusions and risk management recommendations on maize MON 810 remain valid and applicable.

EFSA concludes that, based on the documentation submitted by France, there is no specific scientific evidence, in terms of risk to human and animal health or the environment, that would support the adoption of an emergency measure on the cultivation of maize MON 810 under Article 34 of Regulation (EC) 1829/2003.

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BACKGROUND AS PROVIDED BY THE EUROPEAN COMMISSION AND EFSA

The marketing of maize MON 810 (notification C/F/95/12-02) was authorised under Directive 90/220/EEC in the European Union (EU) for all, other than food, uses by the Commission Decision 98/294/EC of 22 April 1998 (EC, 1998). Consent was granted to the applicant (Monsanto Europe S.A.) by France on 3 August 1998. Food uses of maize derivatives were notified according to Article 5 of the Novel Food Regulation (EC) No 258/97 on 6 February 1998.

On 15 June 2009, the EFSA GMO Panel issued a scientific opinion on the renewal of the authorisation for the continued marketing of: (1) existing food and food ingredients produced from maize MON 810; (2) feed consisting of and/or containing maize MON 810, including the use of seed for cultivation; and (3) food and feed additives, and feed materials produced from maize MON 810. The EFSA GMO Panel concluded that *“maize MON 810 is as safe as its conventional counterpart with respect to potential effects on human and animal health”*, and that *“maize MON 810 is unlikely to have any adverse effect on the environment in the context of its intended uses, especially if appropriate management measures are put in place in order to mitigate possible exposure of non-target (NT) Lepidoptera”*. The EFSA GMO Panel recommended that *“especially in areas of abundance of nontarget Lepidoptera populations, the adoption of the cultivation of maize MON 810 be accompanied by management measures in order to mitigate the possible exposure of these species to maize MON 810 pollen”*. In addition, the EFSA GMO Panel advised that *“resistance management strategies continue to be employed and that the evolution of resistance in lepidopteran target pests continues to be monitored, in order to detect potential changes in resistance levels in pest populations”* (EFSA, 2009).

On 30 November 2011, the EFSA GMO Panel adopted a statement supplementing the environmental risk assessment conclusions and risk management recommendations on maize Bt11 cultivation (EFSA Panel on Genetically Modified Organisms, 2011c). In its statement, the EFSA GMO Panel concluded that *“subject to appropriate management measures, maize Bt11 cultivation is unlikely to raise additional safety concerns for the environment compared to conventional maize”* (EFSA Panel on Genetically Modified Organisms, 2011c). The EFSA GMO Panel considered that the environmental risk assessment conclusions and risk management recommendations on non-target Lepidoptera for maize Bt11 apply equally to maize MON 810 due to the similarities between both *Bt*-maize events (i.e., identity of amino acid sequence of the core of the Cry1Ab protein, similar biological activity against susceptible Lepidoptera, similar Cry1Ab protein expression level in pollen).

The EFSA GMO Panel further supplemented its previous risk management recommendations on maize Bt11 and MON 810 cultivation by reapplying the mathematical model developed by Perry et al. (2010, 2011, 2012), in order to consider additional hypothetical agricultural conditions, and to provide additional information on the factors affecting the insect resistance management (IRM) strategy (EFSA Panel on Genetically Modified Organisms, 2012d).

On 6 December 2012, following a request from the European Commission, the EFSA GMO Panel compiled its previous risk assessment conclusions and risk management recommendations on maize MON 810, and considered their validity in the light of new relevant scientific publications published from 2009 onwards (EFSA Panel on Genetically Modified Organisms, 2012e). Based on the performed literature search, the EFSA GMO Panel concluded that *“its previous risk assessment conclusions on maize MON 810 as well as its recommendations on risk management measures and monitoring remain valid and applicable.”*

Following requests of the European Commission to assess the annual post-market environmental monitoring reports on maize MON 810 cultivation submitted by the applicant, the EFSA GMO Panel issued scientific opinions on the 2009, 2010, 2011 and 2012 PMEM reports on maize MON 810 (EFSA Panel on Genetically Modified Organisms, 2011a, 2012a, 2013c, 2014). The EFSA GMO Panel noted shortcomings in the methodology for case-specific monitoring and general surveillance, and made recommendations to strengthen the annual PMEM activities on maize MON 810. So far, the

data submitted by the applicant in its PMEM reports did not indicate any adverse effects on human and animal health or the environment arising from the cultivation of maize MON 810.

Several EU Member States invoked safeguard clause or emergency measures to provisionally restrict or prohibit the marketing of maize MON 810 on their territory. For all cases for which the EFSA GMO Panel or EFSA has been asked by the European Commission to evaluate whether the invocation was justifiable on the basis of the scientific information submitted in support of a safeguard clause or emergency measures, the EFSA GMO Panel or EFSA concluded that, in terms of risk to human and animal health and the environment, no new scientific evidence had been presented that would invalidate its previous risk assessment conclusions on maize MON 810 (EFSA, 2004, 2005, 2006a, b, 2008a, b, c, d, 2014a; EFSA Panel on Genetically Modified Organisms, 2012b, c, 2013a, b).

TERMS OF REFERENCE AS PROVIDED BY THE EUROPEAN COMMISSION

EFSA is requested in accordance with Article 31 of Regulation (EC) No 178/2002 to provide a statement:

- assessing if the French authorities submitted new scientific evidences in support to their request for a prohibition of GM maize MON 810 cultivation according to Article 34 of Regulation (EC) 1829/2003; and, where appropriate;
- indicating whether these new scientific evidences might lead the GMO Panel to reconsider its previous safety assessments of GM maize MON 810.

ASSESSMENT

At the request of the European Commission, EFSA assessed the documentation supplied by France (referred to hereafter as the French Authorities' report) and the scientific publications cited in the French Authorities' report. For each area of concern outlined in the French Authorities' report, EFSA assessed whether any of the scientific publications not previously addressed by EFSA and/or its GMO Panel, or any of the arguments put forward by France, would invalidate the previous GMO Panel conclusions on the safety of maize MON 810. Moreover, EFSA considered the relevance of concerns raised by the French Authorities in the light of the most recent and relevant scientific data published in the scientific literature.

During its evaluation of the French Authorities' report, EFSA noted that most of the cited scientific publications were addressed previously by EFSA and its GMO Panel in various scientific outputs (e.g., EFSA Panel on Genetically Modified Organisms, 2011b, c, 2012d, 2013c; EFSA, 2014b). These publications are therefore not considered further here, except for HCB (2013), which formed a key part of the basis of the scientific argumentation put forward in the French Authorities' report. In the case of the remaining scientific publications (Campagne et al., 2013; Mezzomo et al., 2013; Zhou et al., 2014), EFSA has focused its assessment on those aspects that are relevant to maize MON 810.

The EFSA assessment below is structured into the Sections used in the French Authorities' report.

1. EFSA assessment of Section I.1, "*Appearance of resistance on target pests*" as referred to in the French Authorities' report

1.1. Cited scientific publications

Of the scientific publications cited in the French Authorities' report with relevance to maize MON 810, one publication (Campagne et al., 2013) was not previously considered by EFSA and/or its GMO Panel. Although the HCB (2013) report was recently considered by the EFSA GMO Panel (EFSA Panel on Genetically Modified Organisms, 2014), it is also addressed here for completeness. The possible relevance of these publications for the risk assessment of maize MON 810 was scrutinised.

- Campagne et al. (2013): The African stem borer (*Busseola fusca*) has evolved high levels of resistance to the Cry1Ab toxin expressed in *Bt*-maize in South Africa. Campagne et al. (2013) investigated the inheritance of Cry1Ab resistance in the African stem borer; they performed controlled crosses with *B. fusca* and evaluated larval survival on Cry1Ab-expressing *Bt*-maize and non-*Bt*-maize. The results showed that survival of resistant larvae and F₁ progeny from resistant × susceptible parents was not lower on Cry1Ab-expressing maize than on non-*Bt*-maize. The authors concluded that: (1) Cry1Ab resistance is inherited dominantly rather than recessively; and (2) insect resistance management (IRM) strategies for Cry1Ab-expressing *Bt*-maize must address the non-recessive inheritance of Cry1Ab resistance in *B. fusca* in South Africa.

- HCB (2013): In the HCB (2013) report, the French Haut Conseil des Biotechnologies (HCB) analysed the susceptibility of the European corn borer (ECB) and Mediterranean corn borer (MCB) (*Ostrinia nubilalis* and *Sesamia nonagrioides*, respectively) to the Cry1Ab toxin over time, using the resistance monitoring data supplied by the applicant (Monsanto, 2013). The HCB considered that the resistance monitoring data indicate that the susceptibility to the Cry1Ab toxin of ECB/MCB populations in Spain is decreased when compared with the reference laboratory strain. Therefore, the HCB recommended: “(1) *re-sampling in 2013 (i.e., prior to 2014) the ECB populations sampled in 2012 in South-West Iberia in order to determine whether there is any increase in their level of resistance to the Cry1Ab toxin; (2) making sure that susceptibility of laboratory reference strains is stable; and (3) improving refuge compliance by MON 810 maize growers in Europe*”.

1.2. Relevance of the cited publications for the risk assessment of maize MON 810

1.2.1. Resistance management (Campagne et al., 2013)

As part of the GMO approval process, applicants submitting a market registration application for cultivation of *Bt*-crops proactively provide an IRM plan (Devos et al., 2013, 2014). IRM plans are designed to reduce the selection pressure associated with *Bt*-crops, in order to prevent or at least delay resistance evolution in the target insect pests (Bates et al., 2005; Alcalde et al., 2007; Andow, 2008; MacIntosh, 2010; Head and Greenplate, 2012). As currently implemented for several *Bt*-crops in several countries, IRM plans usually rely on the high dose/refuge (HDR) strategy (Gould, 1998; Glaser and Matten, 2003; MacIntosh, 2010).

The success of the HDR strategy is aided if the following conditions are met (reviewed by Tabashnik et al., 2013): (1) the *Bt*-toxin is expressed at appropriate levels in relevant plant parts at the appropriate times in relation to the target pest's life cycle; (2) initial resistance alleles are rare in the target insect pest population, so that nearly all resistance alleles will be in heterozygote individuals that cannot survive on the *Bt*-crop; (3) random mating occurs between resistant insects emerging in *Bt*-crops and susceptible insects preserved on refuges at sufficient levels; (4) resistance alleles are partially or fully recessive; and (5) fitness costs are associated with the resistance. Compliance with refuge requirements is an additional critical factor contributing to the success of IRM plans in delaying the rate at which resistance evolves.

When the conditions contributing to the success of the HDR strategy were not met, field-selected resistance to *Bt*-crops occurred, unless refuges were abundant or other (area-wide) integrated pest management (IPM) measures were implemented (reviewed by Tabashnik et al., 2013). Instances of field-selected resistance to *Bt*-maize active against lepidopteran maize pests have been reported in populations of *B. fusca* in South Africa (van Rensburg, 2007; Kruger et al., 2009, 2011; Van den Berg et al., 2013), and in populations of the fall armyworm (*Spodoptera frugiperda*) in Puerto Rico (Matten et al., 2008; Moar et al., 2008; Tabashnik, 2008; Tabashnik et al., 2008a; Storer et al., 2010, 2012), where larvae were able to survive on Cry1Ab-expressing *Bt*-maize MON 810 and Cry1F-expressing *Bt*-maize 1507, respectively.

Reasons for the above-mentioned instances of field-selected resistance range from the autosomally, non-recessive inheritance of resistance to specific agronomic/environmental factors, along with the insufficient planting of refuges of non-*Bt*-maize (Huang et al., 2011; Kruger et al., 2012; Tabashnik et al., 2013; Van den Berg et al., 2013).

EFSA reiterates that the instances of field resistance to lepidopteran-active *Bt*-maize have been reported outside Europe for two target pests in maize that are not present in the European fauna. Moreover, heterozygotes from resistant ECB strains obtained under laboratory conditions did not survive exposure to Cry1Ab-expressing *Bt*-maize plants, supporting the functional recessiveness of resistance to these plants (Siegfried and Hellmich, 2012; Siegfried et al., 2013). If the inheritance of

resistance is not recessive, then modelling results suggest that increasing refuge abundance can still substantially delay resistance (Tabashnik et al., 2008a, b).

At present, EFSA is not aware of early warning signs indicating increases in tolerance to Cry1Ab-expressing *Bt*-maize in field populations of ECB/MCB. Annual assessments of ECB/MCB susceptibility to Cry1Ab in the USA and EU have not revealed any significant change in susceptibility or identified populations that survive on Cry1Ab-expressing *Bt*-maize plants (after more than ten years of exposure to the Cry1Ab protein in the USA) (Farinós et al., 2004, 2011; Stodola et al., 2006; Andreadis et al., 2007; Siegfried et al., 2007; Crespo et al., 2009, 2010; EFSA Panel on Genetically Modified Organisms, 2011a, 2012a, 2013c, 2014; Siegfried and Hellmich, 2012). Moreover, no major resistance alleles have ever been recorded either through laboratory selection experiments (Chaufaux et al., 2001; Siqueira et al., 2004) or by F₂ screening of field populations (Andow et al., 2000; Bourguet et al., 2003; Stodola et al., 2006), strongly suggesting that the frequency of alleles conferring resistance to Cry1Ab-expressing *Bt*-maize plants is <0.001 in all populations examined to date (Siegfried and Hellmich, 2012; Siegfried et al., 2013). The lack of resistance in ECB/MCB and other major insect pests targeted by *Bt*-crops attests that the HDR strategy is capable of preventing, or at least delaying, resistance under field conditions (Andow, 2008; Tabashnik et al., 2008a, b, 2009, 2013; Huang et al., 2011; Siegfried and Hellmich, 2012).

To ensure effective long-term ECB/MCB management and the sustainable use of *Bt*-maize, the EFSA GMO Panel advocated an IPM approach in which *Bt*-maize is only one of many management options. Moreover, resistance monitoring to detect early warning signs indicating resistance evolution in the field, compliance monitoring to assess farmers' compliance with IRM requirements and education (training) programmes aiding farmers to understand the importance of adhering to IRM requirements are essential to the success of the HDR strategy and should therefore continue to form an integral part of IRM plans for *Bt*-maize.

1.2.2. Resistance monitoring (HCB, 2013)

IRM plans for *Bt*-crops require routine monitoring for resistance evolution, so that early warning signs indicating increases in tolerance in the field may be detected (Siegfried and Spencer, 2012). A timely detection of such signs enables actions to limit the survival of resistant insects and to slow or prevent their spread should resistance have evolved among field populations (Siegfried et al., 2007).

Based on the 2012 resistance monitoring data for maize MON 810 (Monsanto, 2013), the applicant concluded that there is no indication of decreased Cry1Ab susceptibility in ECB/MCB populations in Spain. However, using the same dataset, the HCB considered that there is a decrease in susceptibility in the ECB/MCB populations in Spain. The EFSA GMO Panel previously assessed the above-mentioned dataset and the analysis reported by the HCB (2013) (see EFSA Panel on Genetically Modified Organisms, 2014). The EFSA GMO Panel concluded that the potentially increased Cry1Ab tolerance of the ECB/MCB populations, as hypothesised by the HCB, might be due to the declining performance of the reference laboratory strain (e.g., through the infection with pathogens or inbreeding depression). Even though the 2012 dataset showed a trend towards increased values of moulting inhibition concentration (MIC), no significant and consistent decrease in Cry1Ab susceptibility of the ECB and MCB field populations in Spain was observed over time (EFSA Panel on Genetically Modified Organisms, 2014).

In the 2012 PMEM report supplied by the applicant (Monsanto, 2013), the applicant acknowledged that the reference laboratory strain might have shown poor performance, yet the possible reasons were not discussed further. Therefore, the EFSA GMO Panel recommended that the applicant investigates the stability and quality of the reference laboratory strain. The recommendation of the EFSA GMO Panel is consistent with that of the HCB, which advocated "*making sure that susceptibility of laboratory reference strains is stable*".

The susceptibility of the target insect pest has been shown to vary considerably depending upon the source of *Bt*-toxins used (Farinós et al., 2004; Saeglitz et al., 2006). Therefore, it is advisable to use the same *Bt*-toxin source throughout the duration of resistance monitoring. In its 2012 PMEM report, the applicant indicated that it used a new Cry1Ab toxin batch from 2012 onwards, which could have a different biological activity from the initial one used until 2010. The EFSA GMO Panel therefore assessed whether the hypothesised change in susceptibility could be attributed to the use of different Cry1Ab toxin batches. A study, performed by the applicant, indicated that the initial and new *Bt*-toxin batch have similar biological activity on ECB (Appendix 8 in Monsanto 2013). It is therefore considered unlikely that the hypothesised change in susceptibility can be attributed to the use of different Cry1Ab toxin batches.

To ensure an early detection of change in susceptibility of the ECB and MCB field populations, the EFSA GMO Panel strongly reiterated its previous recommendation for annual sampling of ECB/MCB in areas of high maize MON 810 adoption rate, especially in north-east Spain in 2014 (EFSA Panel on Genetically Modified Organisms, 2013d).

1.3. EFSA conclusion

Neither the results reported by Campagne et al. (2013) and HCB (2013) nor the arguments put forward by France in the French Authorities' report reveal any new information that would invalidate the previous risk assessment conclusions and risk management recommendations made by the EFSA GMO Panel.

2. EFSA assessment of Section I.3, "*Impact of MON810 maize on non-target invertebrates*" as referred to in the French Authorities' report

2.1. Cited scientific publications

From the scientific publications in the French Authorities' report with relevance to maize MON 810, two publications (Mezzomo et al., 2013; Zhou et al., 2014) were not previously considered by EFSA and/or its GMO Panel. The possible relevance of these publications for the risk assessment of maize MON 810 was scrutinised.

- Mezzomo et al. (2013): Mezzomo et al. (2013) studied the potential haematotoxicity (by conventional haematology) and genotoxicity (by *in vivo* micronucleus test) of spores of four *Bt*-strains expressing Cry toxins in Swiss albino mice. These *Bt*-strains were genetically modified to express Cry1Aa, Cry1Ab, Cry1Ac or Cry2Aa. The spores, re-suspended in distilled water, were given by oral gavage in a single administration of one of three doses (27, 136 or 270 mg of lyophilised spores/kg body weight (bw)). Binary combinations of lyophilised spores were also given at 270 mg/kg bw. The negative control group received distilled water (vehicle). As a positive control for the micronucleus test, mice received cyclophosphamide at 27 mg/kg bw. Group size was three male and three female mice. Blood and bone marrow were sampled after 24 hours in all groups. Blood samples were also taken after 72 hours or 7 days from additional groups receiving the high dose of the individual *Bt*-strains or 27 mg cyclophosphamide/kg bw. Based on the results, the authors concluded that administrations of *Bt*-spores provoked selective haematotoxicity, particularly for the erythroid lineage; a significant reduction in bone marrow cell proliferation demonstrated cytotoxic but not genotoxic effects.
- Zhou et al. (2014): Zhou et al. (2014) reported that the Cry1Ab toxin affects the metabolic enzymes acetylcholine esterase, glutathione peroxidase and superoxide dismutase in the predatory spiders *Ummeliata insecticeps* and *Pardosa pseudoannulata* when fed Cry1Ab-containing prey (fruit flies). The authors concluded that plant-produced *Bt*-proteins can affect non-target arthropods at the physiological and biochemical level, and reduce their fitness.

2.2. Relevance of the cited publications for the risk assessment of maize MON 810

2.2.1. Haematotoxicity (Mezzomo et al., 2013)

The test items were not purified Cry toxins, but viable *Bt*-spores expressing the Cry toxins. In addition, the *Bacillus* species used is known to be capable of producing enterotoxins (e.g., Gavidia Rivera et al., 2000) and a range of other toxic materials (Butko, 2003). Therefore, in the absence of an appropriate *Bt*-control strain, it is not possible to attribute any observed findings specifically to the Cry toxins, as was done by Mezzomo et al. (2013) in describing their results.

This acute study showed several key weaknesses which impede interpretation of the results. First, a study which addresses only peripheral blood findings of haematotoxicity has limited value in the absence of gross morphological and/or histopathological investigations. Haematological changes may be secondary to other pathological conditions (for example to haemorrhages) which were not monitored in this study. Second, vehicle control groups for the evaluation of peripheral blood analyses at 72 hours and 7 days after administration were missing. Finally, there is a lack of data indicating normal variation of the measured parameters under the study conditions, particularly considering the small number of animals per sex and per group and the use of a single vehicle control group (at 24 hours after administration). In addition, the micronucleus test was not performed in accordance with the current standards (OECD Guideline TG 474), in particular with respect to the number of animals used (only three animals per sex and group, instead of five), the dosing regime (sampling at only one time point, instead of at least two), as well as the presentation and interpretation of results (only group data but not individual data were considered).

Regarding the endpoints measured 24 hours after administration, the authors reported that four parameters of the erythrogram (mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), mean corpuscular volume (MCV) and red cell distribution width (RDW)) showed statistically significant differences compared with the vehicle control group, but no significant differences were identified in the leucogram. Overall, EFSA considers that the values of the erythrogram parameters for the four *Bt*-strains and for the three dose levels are very similar. The only notable difference remained between the test groups and vehicle control group for the MCV and its coefficient of variation (RDW). These differences were small in magnitude (around 10 %) and were not dose-related. Furthermore, these parameters are derived by calculation from the haematocrit, red blood cells (RBCs) and haemoglobin content of the blood, none of which showed a significant difference. Decreases in the percentage of polychromatic erythrocytes in the bone marrow of treated animals relative to the vehicle control group were also used as an argument to support effects on the erythroid lineage by the authors. However, owing to the limitations of the micronucleus test performed, this conclusion is not justified.

The authors also reported significant differences in platelet counts and related parameters (mean platelet volume (MPV), platelet large cell ratio (P-LCR) and platelet distribution width (PDW)) measured 24 hours after administration. Platelet counts are known to be highly variable, and the corresponding figures for the three dose levels showed no evidence of dose-effect relationship.

Based on these findings, EFSA concludes that the Mezzomo et al. (2013) publication does not support the conclusions on haematotoxicity associated with the Cry toxins.

2.2.2. Potential adverse effects on spiders (Zhou et al., 2014)

The findings reported by Zhou et al. (2014) suggest that the Cry1Ab toxin can affect enzyme activity in predatory spiders when fed Cry1Ab-containing prey. Even though the spiders have been shown to take up the Cry1Ab toxin when feeding on the Cry1Ab-containing prey, no correlation can be made between the Cry1Ab toxin content measured in the spiders and the observed differences in enzyme activity in the spiders. Some limitations of the study (such as undefined number of organisms tested

and replications; lack of details on how the protoxin was quantified in the diet; ill-characterised and -described purity of the Cry1Ab toxin; lack of a positive control) make it difficult to explain the mechanism leading to the observed differences. Therefore, EFSA considers that the dataset reported by Zhou et al. (2014) is preliminary and is not sufficient to demonstrate a new mode of action of the Cry1Ab protein on spiders.

In addition, the biological relevance of the observations made remains difficult to assess, as it is not clear how the observed differences in enzyme activity are related to fitness parameters of the spiders; the authors did not report on life-table parameters of the spiders. Moreover, the reported response was transient.

Finally, exposure has not been sufficiently characterised. The test model used by Zhou et al. (2014) does not reflect realistic exposure conditions; spiders are generalist predators, and the level of exposure to the Cry1Ab toxin will depend on the prey spectrum of individual species. In the experiments performed by Zhou et al. (2014), the variability of food uptake was not controlled, and this may have influenced the observed variability of the Cry1Ab toxin content in the spiders. Therefore, it remains challenging to extrapolate the results of the observations made under controlled conditions to spider activity under field conditions.

At present, EFSA is not aware of identified significant adverse effects of the Cry1Ab toxin on spiders. Laboratory studies have indicated that plant-produced Cry proteins have no direct effects on life-table parameters of spiders after ingestion, whereas field studies confirmed that population densities of spiders are not adversely affected (reviewed by Peterson et al., 2011). Those findings are in line with the outcomes of meta-analyses, in which a broad range of beneficial arthropods including spiders were addressed (Marvier et al., 2007; Wolfenbarger et al., 2008; Naranjo, 2009; Peterson et al., 2011; Albajes et al., 2013; Comas et al., 2014).

2.3. EFSA conclusion

Neither the results reported by Mezzomo et al. (2013) and Zhou et al. (2014) nor the arguments put forward by France in the French Authorities' report reveal any new information that would invalidate the previous risk assessment conclusions and risk management recommendations made by the EFSA GMO Panel.

3. EFSA assessment of Section II, “*Management measures needed to protect the environment*” as referred to in the French Authorities' report

Section II of the French Authorities' report lists all the recommendations for risk management measures made by the EFSA GMO Panel in of its various scientific outputs (e.g., EFSA, 2009; EFSA Panel on Genetically Modified Organisms, 2011a, c, 2012a, d, e, 2013c, 2014), and does not present any new scientific information.

4. EFSA assessment of Section III, “*Insufficient implementation of management measures*” as referred to in the French Authorities' report

Section III of the French Authorities' report focuses on the legal enforcement of EFSA's recommendations. This issue is not in EFSA's remit and therefore not considered further.

5. Conclusions

Neither the scientific publications cited in the French Authorities' report with relevance to maize MON 810 nor the arguments put forward by France reveal any new information that would invalidate the previous risk assessment conclusions and risk management recommendations made by the EFSA GMO Panel. Therefore, EFSA considers that the previous GMO Panel risk assessment conclusions and risk management recommendations on maize MON 810 remain valid and applicable.

EFSA concludes that, based on the documentation submitted by France, there is no specific scientific evidence, in terms of risk to human and animal health or the environment, that would support the adoption of an emergency measure on the cultivation of maize MON 810 under Article 34 of Regulation (EC) 1829/2003.

DOCUMENTATION PROVIDED TO EFSA

1. Letter from the European Commission, dated 25 March 2014, to the EFSA Executive Director requesting the assessment by EFSA of the scientific elements supporting the French request for a prohibition of the placing on the market of GM maize MON 810 for cultivation purposes in the EU.
2. Acknowledgement letter, dated 16 April 2014, from the EFSA Executive Director to the European Commission.

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